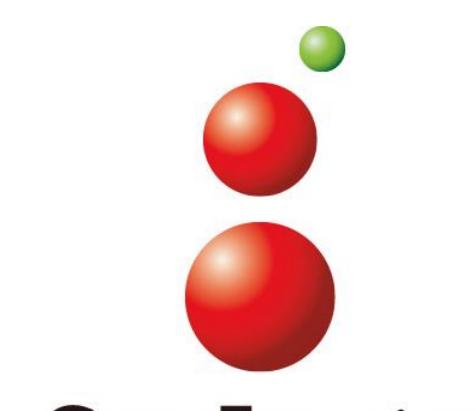


PURE systemにおけるアンバーアプレッションによる非天然アミノ酸導入の最適条件の探索

Exploration of optimal conditions for the incorporation of noncanonical amino acids by amber suppression using the PURE system

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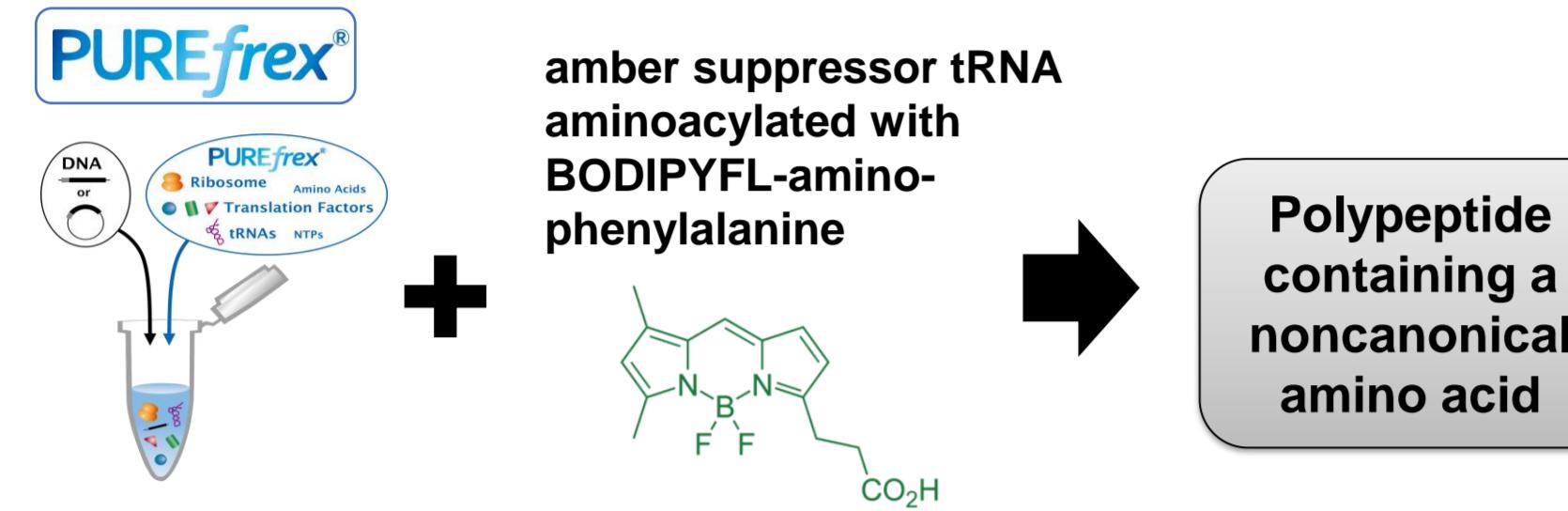
° Rena Matsumoto, Takayoshi Watanabe, Takahiro Hohsaka and Takashi Kanamori (¹GeneFrontier Corp., ²School of Material Science, JAIST)



<Abstract>

PUREflex®は、PURE systemを基にした大腸菌でのタンパク質合成に関与する因子のみから再構成した無細胞タンパク質合成系である。反応液組成の調節が容易なため、サプレッサーtRNAに非天然アミノ酸を結合させ、終止コードン(アンバーコードン(UAG))に非天然アミノ酸を導入するアンバーアプレッションにも適している。しかし、これまでPURE systemを用いたアンバーアプレッションによる非天然アミノ酸の導入において、反応液の構成因子についての詳細な条件検討は行われていなかった。そこで、我々はPUREflex®を用いたアンバーアプレッションの最適条件を探索した。モデルタンパク質として大腸菌ジヒドロ葉酸還元酵素(DHFR)のN末端側にS-tag (KETAAAKFERSQHMDS)を付加したコンストラクトを行い、BODIPYFL-amino-phenylalanine (BFLAF)が付加されたアンバーサプレッサーtRNAによりS-tag中にBFLAFが導入されたタンパク質を合成し、サプレッション効率を比較した。はじめにS-tag中のアンバーコードン最適位置を検討したところ、S-tag中のT7、F12の位置が最もサプレッション効率が高かった。これらのコンストラクトを行い、BFLAF-サプレッサーtRNA添加量や反応液中の構成因子の濃度を検討した。その結果、効率的なアンバーアプレッションには、競合する翻訳終結因子RF1を反応液中から除くことが必要とされていたが、実際には一定濃度のRF1を添加した反応液を用いるとサプレッション効率が高くなることが分かった。また、アンバーコードンの1つ前のコドンの違いでサプレッション効率および全長の合成産物が得られる効率が異なることも分かった。

Incorporation of noncanonical amino acids by amber suppression using PUREflex®



Purpose of this study

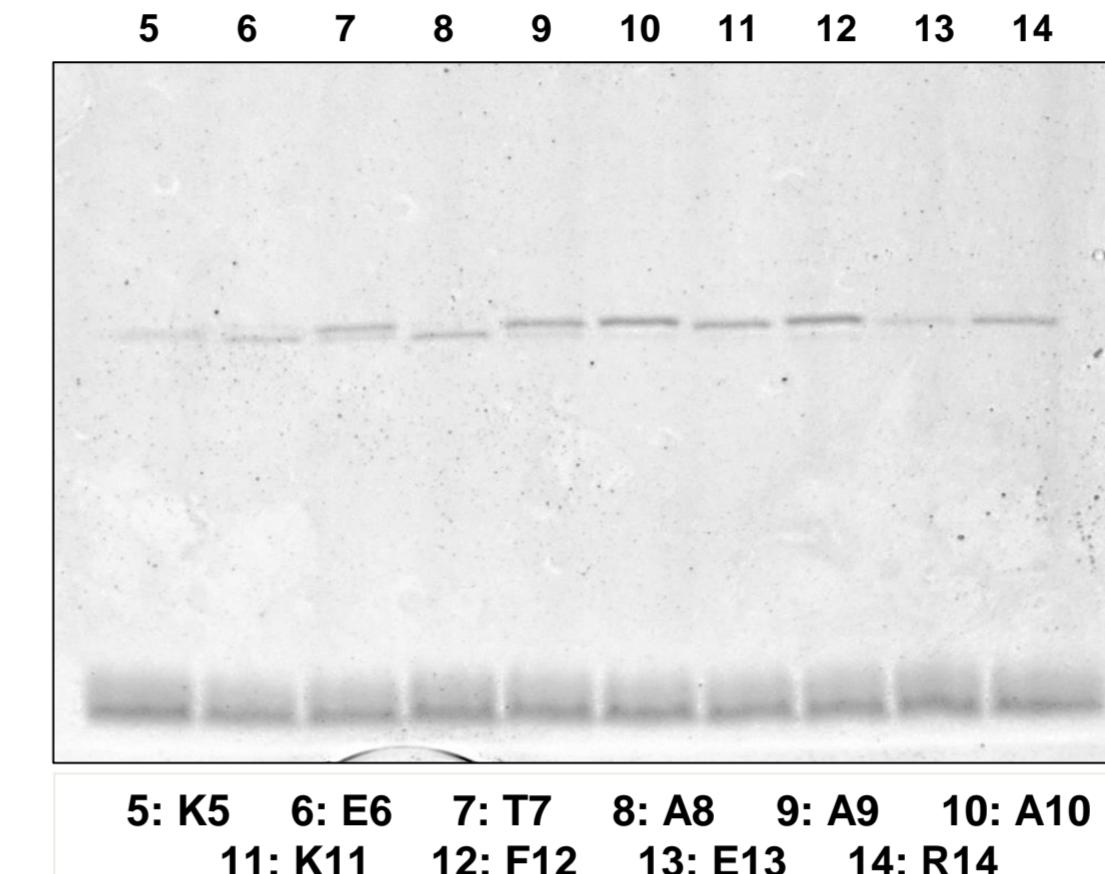
Exploration of the optimal conditions for incorporation of a noncanonical amino acid into the N-terminal region by amber suppression

1. Optimal position of amber codon for incorporation of a noncanonical amino acid

Model protein: Stag-DHFR-FLAG

MNKL KETAAAKFERSQHMDS DHFR FLAG

Stag-DHFR-FLAG 1 ATGAAATAGCTTAAAGAAGAGTCGTAATAATTGAAAGTCAACATATGGATTCATC 60
M N K L K E T A A A K F E R Q H M D S I
K5am 1 ATGAAATAGCTTAAAGAAGAGTCGTAATAATTGAAAGTCAACATATGGATTCATC 60
M N K L K E T A A A K F E R Q H M D S I
E6am 1 ATGAAATAGCTTAAATAGTCGTCAGGTAAATAATTGAAAGTCAACATATGGATTCATC 60
M N K L K E T A A A K F E R Q H M D S I
T7am 1 ATGAAATAGCTTAAAGAAGAGTCGTCAGGTAAATAATTGAAAGTCAACATATGGATTCATC 60
M N K L K E T A A A K F E R Q H M D S I
A8am 1 ATGAAATAGCTTAAAGAAGAGTCGTCAGGTAAATAATTGAAAGTCAACATATGGATTCATC 60
M N K L K E T A A A K F E R Q H M D S I
A9am 1 ATGAAATAGCTTAAAGAAGAGTCGTCAGGTAAATAATTGAAAGTCAACATATGGATTCATC 60
M N K L K E T A A A K F E R Q H M D S I
A10am 1 ATGAAATAGCTTAAAGAAGAGTCGTCAGGTAAATAATTGAAAGTCAACATATGGATTCATC 60
M N K L K E T A A A K F E R Q H M D S I
K11am 1 ATGAAATAGCTTAAAGAAGAGTCGTCAGGTAAATAATTGAAAGTCAACATATGGATTCATC 60
M N K L K E T A A A K F E R Q H M D S I
F12am 1 ATGAAATAGCTTAAAGAAGAGTCGTCAGGTAAATAATTGAAAGTCAACATATGGATTCATC 60
M N K L K E T A A A K F E R Q H M D S I
E13am 1 ATGAAATAGCTTAAAGAAGAGTCGTCAGGTAAATAATTGAAAGTCAACATATGGATTCATC 60
M N K L K E T A A A K F E R Q H M D S I
R14am 1 ATGAAATAGCTTAAAGAAGAGTCGTCAGGTAAATAATTGAAAGTCAACATATGGATTCATC 60
M N K L K E T A A A K F E R Q H M D S I



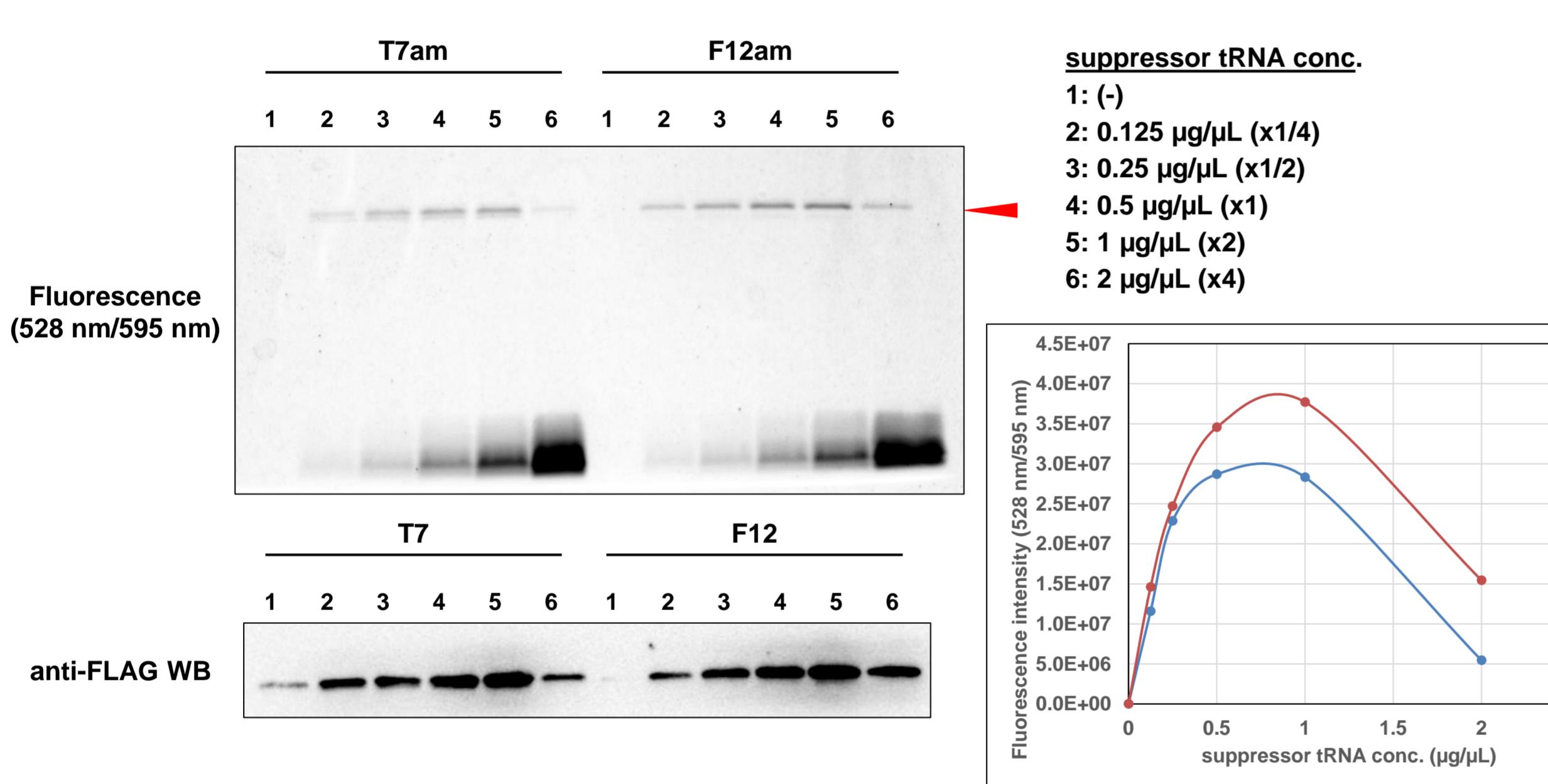
ver. 1 (ΔRF1)
+ 0.5 µg/µL amber suppressor tRNA
aminoacylated with BODIPYFL-
amino-phenylalanine (BFLAF-
tRNAcua)
1 ng/µL Stag(am)-DHFR-FLAG
reaction at 37 °C for 60 min

The amount of the product containing BODIPY was higher at positions T7, A10 and F12, than at other positions. However, the efficiency of the incorporation of BFLAF was low.

2. Optimization of the concentration of three factors in the reagent for amber suppression

2-1. suppressor tRNA

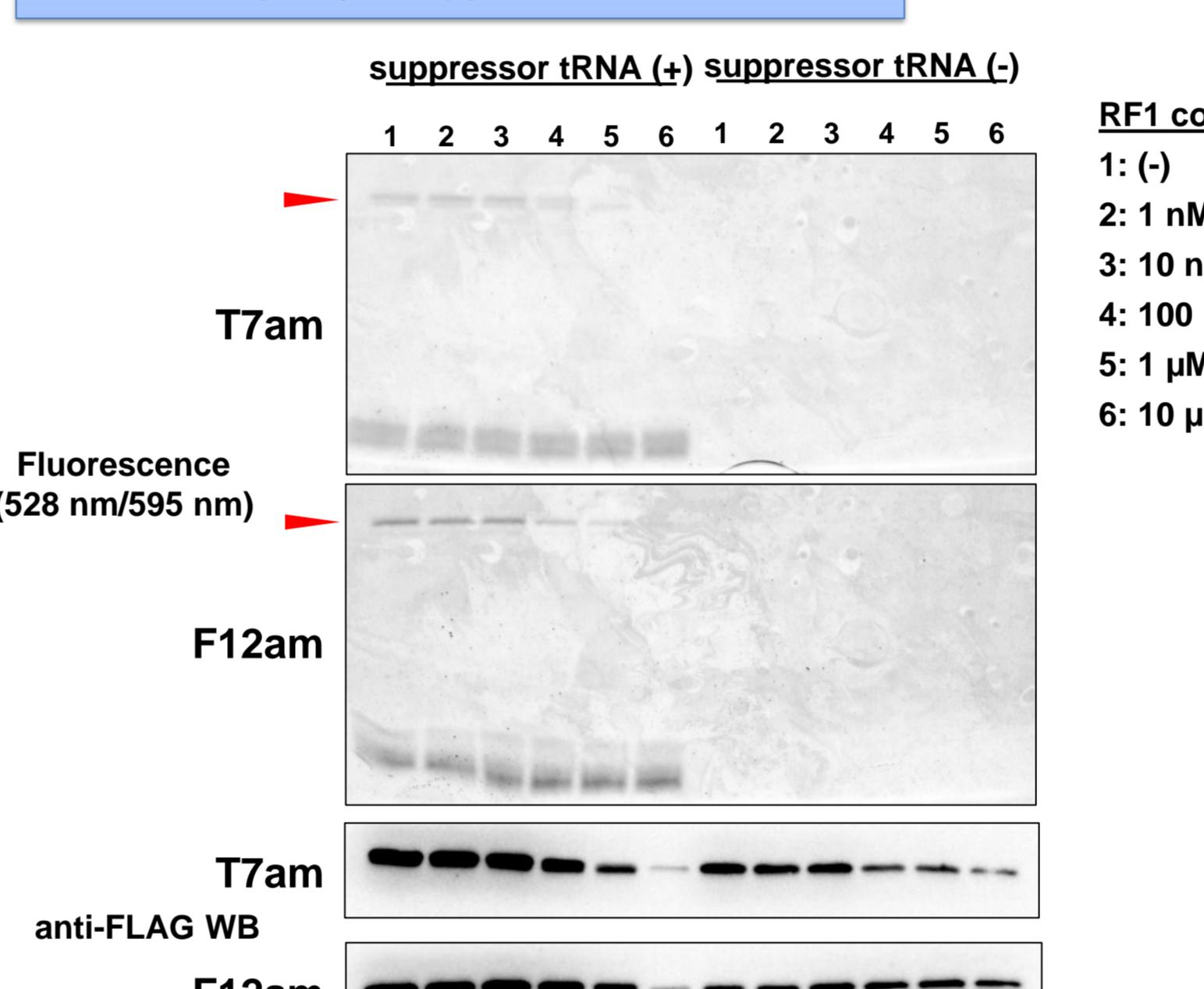
ver. 1 (ΔRF1)
+ 0-2 µg/µL BFLAF-tRNAcua
1 ng/µL Stag(T7am or F12am)-DHFR-FLAG
reaction at 37 °C for 60 min



The product containing BFLAF exhibited high expression levels when 0.5 or 1 µg/µL of suppressor tRNA was applied. However, the productivity was decreased when 2 µg/µL suppressor tRNA was used.

2-2. RF1

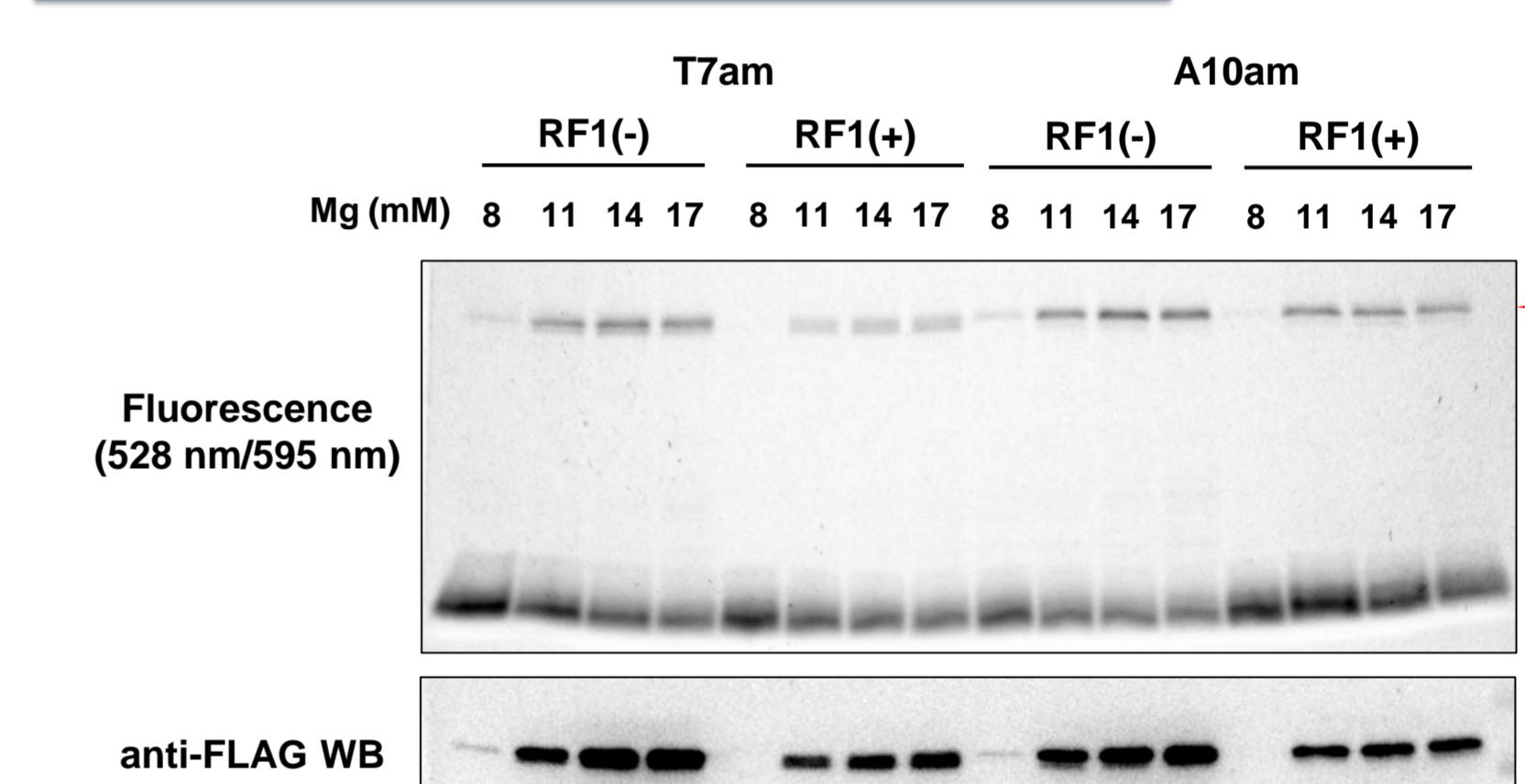
ver. 1 (ΔRF1)
+ 0/0.25 µg/µL BFLAF-tRNAcua
+ 0-10 µM RF1
1 ng/µL Stag(T7am or F12am)-DHFR-FLAG
reaction at 37 °C for 60 min



The suppression efficiency was comparable to that observed in the absence of RF1, even at concentrations below 100 nM. Additionally, full-length products were synthesized without using suppressor tRNA.

2-3. Magnesium

ver. 1 (ΔRF1/ΔCTP/ΔUTP/ΔT7RNAP)
+ 0.25 µg/µL BFLAF-tRNAcua
+ 0/0.1 µM RF1
Mg: 8-17 mM
0.2 µM Stag(T7am or A10)-DHFR-FLAG mRNA
reaction at 37 °C for 60 min



The product containing BFLAF was synthesized with high efficiency at 14 mM magnesium or higher.

sup. tRNA: 0.5-1 µg/µL
RF1: 0-100 nM
Magnesium: > 14 mM

3. Optimal codon immediately before amber codon

In *E. coli* S30-extract system, it has been shown that the codon immediately preceding amber codon affects the efficiency of amber suppression using BFLAF.



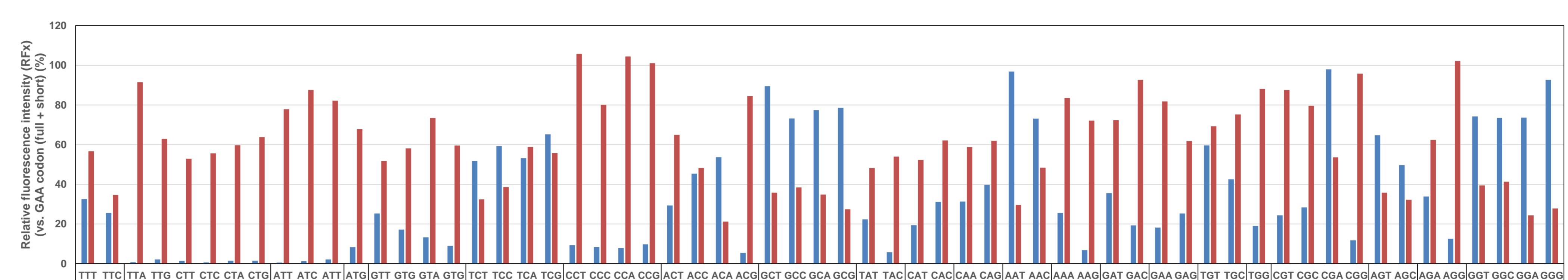
Construction of DNA in which the codon immediately preceding amber codon was substituted

Stag (X6T7am)-DHFR-FLAG

MNKL KX*AAAKFERSQHMDS DHFR FLAG

X=F (ttt)
F (ttc)
L (tta)
L (tgt)
.....
* = tag (amber codon)

F	TTC	TTA	TTG	CTT	CTG	CTA	CTG	I	M	V	TCC	TCA	TCG	CCT	CCC	CCA	P	T	A	G	Y	H	Q	N	K	D	E	C	W	R	S	R	G
Fluorescence (528 nm/595 nm)																																	



Why is the short-length product synthesized?

full-length MNKL KX*AAAKFERSQHMDS DHFR FLAG

short-length *AAAKFERSQHMDS DHFR FLAG

It is hypothesized that a product with an N-terminal amber codon is synthesized.

The codon immediately upstream of amber codon significantly affected the synthesis by amber suppression using PUREflex.

Good codon: Ala (GCT/ GCC/ GCA/ GCG)
Gly (GGT/ GGC/ GGA/ GGG)
Arg (CGA)
Asn (AAT)

4. Conclusion

- A noncanonical amino acid was incorporated by amber suppression even in the presence of RF1 (or it could be better with RF1).
- The position of amber codon and a codon immediately before amber codon significantly affected the suppression efficiency.

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